# **Determination the Inhibition activity of Magnesium Oxide Nanoparticles Synthesized by** *Aspergillus niger* **against some uncommon bacterial species which isolated from different sources of infection**

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## **Abstract**

The resistance of bacteria to chemically manufactured antibiotics is dangerous in human health. this has made the need for continuous detection of diverse resources ,especially from natural sources in the treatment of human diseases. The current study employs *Aspergillus niger* for the formation of of Magnesium Oxide Nanoparticles, and it involves the antimicrobial activity of the particles against some of uncommon bacterial species which include Gram positive *Aeromonas salmonicida, Kocuria rosea and Alloiococcus otitis* and Gram negative *Acinatobacter calcoaceticus, Pantoea agglomerans* and *Ochrobactrum anthropic* by agar wells diffusion method. The results were indicated that biosynthesized of MgO NPs were found as extracellular and it was appear in the range of size 40-90 nm through characterization technique such as Scanning Electron Microscopy (SEM), and assurance by UV-Visible spectroscopy for the absorbance band at 258 nm, The results revealed that MgO NPs is an effective antibacterial agent against Gram positive *Aeromonas salmonicida , Kocuria rosea and Alloiococcus otitis* and the zone inhibition diameter was at 20,19 and23 mm respectively compared with inhibition effects of ciprofloxacin antibiotics at 18, 17and 20 mm respectively. while against Gram negative *Acinatobacter calcoaceticus, Pantoea agglomerans* and *Ochrobactrum anthropic* bacteria was at 18,20,19 mm respectively, compared with inhibition effects of ciprofloxacin antibiotics at 16, 20 and 15 mm respectively.

*Keywords: Nanoparticles, Biosynthesis, Aspergillus niger, Antibacterial activity, Uncommon bacteria.*

**تحذيذ الفعبلية التثبيطية لجزيئبت اوكسيذ المغنسيوم النبنوية المصنعة** *niger Aspergillus* **بواسطة الفطرضذ بعض انواع البكتريب غير الشبئعة والمعزولة من مصبدر اخمبج مختلفة**

# **دمحم نظير معروف**

**الملخص**

إن مقاومـــة البكتيريــــا للمضــــادات الحيويـــة المصـــنعة كيميائيـــا تشـــكل خطـــر ا علـــي صـــحة الإنســـان، ممــــا جعـــل الحاجــة إلـــم الكشّــف المســتمر عــن المــوارد المتنوعــة، وخاصـــة مــن المصـــادر الطبيعيــة فـــى عــلاج امــراض الانســــان. اعتمــــدت الدراســــة الحاليــــة علــــى تصـــــنيع جزيئــــات اوكســـيد المغنســـيوم النانويــــة بواســــطة الفطــــر *niger Aspergillus* وتحايةةةةةا ف اليتثةةةةةا الت بيطيةةةةة ةةةةةا ةةةةة األوةةةةةوا البكتيريةةةةة غيةةةةةر المألوفةةةةة والم سولةةةةة مةةةةةةةه مصةةةةةةةا اخمةةةةةةةا م تل ةةةةةةة والتةةةةةةة وةةةةةةةملل اوةةةةةةةوا موجبةةةةةةة لصةةةةةةةب كةةةةةةةرا وهةةةةةةة *Aeromonas otitis ام*ا الانواع السالبة لصبغة كرام فهي salmonicida , Kocuria rosea , Alloiococcus otitis *Ochrobactrum* و*Acinatobacter calcoaceticus, Pantoea agglomerans*

anthropic المقتدام طريقة الانتشـــار فــــي الحفـــر، وتميـــزت النتــــائج بقـــدرة الفطــــر Aspergillus niger علمي تصميني جزيئمات المغنسميوم النانويسة وباحجسام تتسراوح بسين 40-90 نسانومتر مسن خسلال .<br>الكشَّــف عـــن خصائصــــها النانويــــة بواســـطة تقنيـــة المجهـــر الإلكترونــــيّ الماســـح، والامتصاصـــية عنـــد طـــول مـــــوجي 258 نـــــانومتر باســـــتخدام جهـــــاز Spectroscopy- - Spectroscopy، واظهـــــرت نتـــــائج الفعاليـــــة التثبيطيـــة لجزيئــــات اوكســـيد المغنســـيوم النانونيــــة تأثير هــــا الفعــــال ضــــد انــــواع البكتريــــا الموجبــــة لصــــبغة بةةةةي ت اقطةةةةا *Aeromonas salmonicida , Kocuria rosea , Alloiococcus otitis* كةةةةرا 03,20 و23 ملــــم علـــــى التـــــوالي مقارنــــــة مـــــع التـــــاثير التثبيطــــي لمضـــــاد Ciprofloxacin والـــــذي يعتبـــــر 01,02 و84 وةةةةةةةاوومتر علةةةةةةة التةةةةةةةوال , يىمةةةةةةةا ةةةةةةةا االوةةةةةةةوا السةةةةةةةالب لصةةةةةةةب كةةةةةةةرا *Acinatobacter* اقطةةةةةةةا فكاوةةةةةةةل *Ochrobactrum anthropi*c<sup>و</sup> *calcoaceticus, Pantoea agglomerans* التثبــــــيط 18,02و19 نـــــــانومتر علـــــــى التــــــوالي مقارنـــــــة مـــُـــع اقطـــــار تثبــــــيط مضــــــاد Ciprofloxacin 20,16و15 على التوال*ي .* الكلمـــــات المفتاحيــــــة :الجزيئــــــات النـانويــــــة ،البنــــــاء الحيـــــو ي ، Aspergillus niger ،الفعاليــــــة التثبيطيـــــة **للبكتريب و البكتريب الغير شبئعة .**

## **Introduction**

The metallic nanoparticles are the most promising material as antibacterial activity, and it gain the current interest in research due to the growing microbial resistance against antibiotics and the developing of the resistant strain (1). Nanotechnology is a field that is developing day by day, making an impact in all spheres of human life and creating a growing sense of excitement in the life sciences especially biomedical devices and biotechnology (2). At the nanoscale level, materials have distinct chemical, physical, optical, magnetic and electrical properties due to their large surface area to volume ratio (3). one of the most important aspects of nanotechnology is the synthesis of nanoparticles (NPs), which form the essence of the nanomaterials (4). Nanoparticles exhibit new pr0perties based 0n specific characteristics such as size, distribution, and morphology (5). Nowadays, nanoparticles are used in many fields including manufacturing and materials, the environment, energy and electronics and in medicine. MDR is a growing problem in the treatment of infectious diseases due to the widespread use of broad spectrum antibiotics has resulted in produced antibiotic resistance for many human bacterial pathogens (6).

correlate the biological activity of inorganic antibacterial agents with the size of the constituent particles (7, 8). In particular, inorganic oxide nanomaterials like Cao, Zno and Mgo have shown potential as effective alternatives in addressing some of these challenges (9). Mgo NPs have the advantage of non-toxicity, high thermal stability, biocompatible, low cost, and have considerable potential as an antibacterial agent. Magnesiume plays several vital roles in human biology (10). The mechanism of metal oxide nanoparticle acti0n 0n bacteria is c0mplicated and n0t fully underst00d. It has been rep0rted that the antibacterial activity of Mgo nanoparticles is attributed to the production of reactive oxygen species (RoS) which induce lipid peroxidation in bacteria (11). Several studies have shown that smaller particles have greater antibacterial activity due to higher reactive surface area (12). Biological methods for nanoparticle synthesis using microorganisms, enzymes, and plants or plant extracts have been suggested as possible ecofriendly alternatives to chemical and physical methods (13, 14). Therefore this study aimed to synthesize Mgo NPs by biological method and characterize the synthesized NPs by utilizing UV-vis, TEM. Besides, their antibacterial activity against

Many researchers have attempted to

some uncommon bacteria were tested. (Gram positive *Aeromonas salmonicida, Kocuria rosea and Alloiococcus otitis* and Gram negative *Acinatobacter calcoaceticus, Pantoea agglomerans* and *ochrobactrum anthropic*) used well diffusion method.

## **Materials and methods Biosynthesis of Mgo nanoparticles**

The Mgo NPs were prepared using method 0f (15). with some modification. The synthesis of Mgo NPs was carried with *Aspergillus. niger* ATCC 16404, the active culture of the isolate was inoculated into Potato dextrose broth (PDB) and the flasks were incubated at 28 °C  $\pm$ 2, 150 rpm for 3 days. After incubation, the fungal filtrate was obtained by passing through Whatman No.1filterpaper. The collected supernatant was added to deionized water treated with 1% 0f mM MgCL2 and further incubated with shaker incubator at 150 rpm for 96 hrs at 28 °C. Conical flasks with either fungal filtrate or MgCL2 served as positive and negative control respectively.

## **UV-vis spectra analysis**

The UV-Visible spectra of Mgo nanoparticles were characterized UV VIS spectrophotometer measured using a Systronics UV double-beam spectrophotometer. This is a simple method that give information about particle concentration and size, and size/size distribution, the shift of absorbance relay on the size (diameters) and shape of particles. The scanning range for the samples was 200-600 nm at a scan speed 0f 480 nm/min. The spectrophotometer was equipped with "UVWinlab" software to record and analyze data. Base line correction of the spectrophotometer was carried out by using a blank reference (16).

## **Scanning electron microscope (SEM)**

A scanning electron microscope was (Cam Scan-3200 LV SEM machine) used to

record the micrograph images, characterize mean particle size and morphology of synthesized Mgo NPs. A thin layer of gold was coated in an auto fine coater to make the samples conductive, after that the material was subjected to analysis by SEM machine was operated at a vacuum (14 , 17).

## **Transmission electron microscope (TEM)**

The shape and size of silver nanoparticles were determined by TEM. For TEM, a drop of Mgo NPs suspension was placed on a carbon-coated copper grid then the grid was dried by Whatman paper No. 1 for 30 minutes. The carbon-coated grid was set on Phillps CM10 TEM sample holder. The average size and shape of Ag NPs was determined at magnification of 130.000×, 80 kV Modified procedure from (18)

## **Antimicrobial activity of synthesized Mgo nanoparticles**

All pathogenic uncommom bacteria were multidrug resistant; they have obtained from out and in patients admitted in Tikrit General Hospital. The collected clinical strains were isolated from urine and sputum specimens. identification of pathogenic bacterial isolates according to the standard methods which recommended by (19,20). Confirmed by VITEK 2 System. The antibacterial efficacy of the Mgo NPs was investigated by agar well diffusion  $assav(21)$ . against various types of multidrug resistant bacteria isolated from urine samples. The tested uncommon bacteria included (Gram positive *Aeromonas salmonicida , Kocuria rosea and Alloiococcus otitis* and Gram negative *Acinatobacter calcoaceticus, Pantoea agglomerans* and *0chr0bactrum anthropic*) Approximately 20ml of Mueller Hinton agar media was poured in sterilized petri dishes. Fresh overnight cultures of pathogenic bacterial isolates were adjusted  $1.5 \times 10^8$ cells/ml, matching with 0.5 McFarland as

mentioned previously . Inoculums (100μl) were applied on the surface of the Muller Hinton agar plates and spread by using swabbed onto the plates. Agar wells of 5 mm diameter were prepared with the sterilized cork borer. Three wells were bored, one well containing the extract alone, control positive and the other well loaded with the synthesized Mgo NPs. The well added of Mgo- NPs 100 μg/ml and 0.50 mg/ml Ciprofloxacin antibiotics used as positive control against bacterial isolates. Then the plates were incubated at 37˚C for 24 hrs, where upon inhibitory activity was observed as a zone of clearing around the wells. The diameter of the clearing zones was measured in mm using the ruler scale. Pathogenic bacterium and compared with the standard antibiotic (22, 23).

# **Results and Discussion**

## **Biosynthesis of Mgo nanoparticles:**

The biosynthesis of Mgo NPs was conducted by cultivation of *Aspergillus niger* on potato dextrose broth media enrichments with ZnCL2 at 28  $^{\circ}$ C  $\pm$ 2with agitation at 150 rpm for 72 hrs., The Mgo NPs extracellular synthesis by cultivati0n of *Aspergillus niger* ATCC 16404 mycelia

biomass, on optimal media enrichment with  $MgCL<sub>2</sub>$ , The quantity of Mgo NPs was at 56 mg/100 ml which were collected after centrfuge and drying on 60 °C. In the biosynthesis of metal oxide nanoparticle by a fungus, the enzymes in metabolic pathway are action in reduce a metal to its metallic solid nanoparticles through the catalytic effects (24, 15). Many questions remain unanswered about the resistance mechanism of fungal strains towards many inhibitors such as heavy metals. Although fungi possess many properties that influence metal toxicity, the mechanisms involved in metal tolerance are highly dependent on the metabolic and nutritional status of the organism (25)

## **Characterization of Mgo NPs**

The prepared of Mgo NPs was characterized using UV-VIS spectroscopy. Fig.1. and the absorption was appearing as a band at 258 nm. These results were agreement with the UV-visible spectrum for Mgo NPs at 260 nm in the results 0f (26). The optical properties of the Mgo NPs were studied in detail by means of the UV–vis absorption spectra in the wavelength range of 200–600 nm at room temperature.





The determine of purity and the size range of Mgo NPs biosynthesized with *A. niger* used Scanning electron microscope analysis was appeared the size range from 47.35 to 98.46 nm (Fig.3). These results were agreement with the results obtained by (27).

the biosynthesis of nanoparticles were induced the microbial cells to produced the biological agents that secrete a large amount of enzymes, which are capable of hydrolyzing metals, and produced the metals ions (28).



**Fig. (2):- The SEM images of the of Mgo NPs.**

### **Isolation and Identification of bacterial isolates**

The isolated uncommon of bacteria from urine and sputum source, which were identified according to the microscopic, macroscopic and biochemical tests and the results confirmed by VITEK-2 SYSTEM. The most frequently isolated bacteria were (Gram positive *Aeromonas salmonicida, Kocuria rosea and Alloiococcus otitis* and Gram negative *Acinatobacter calcoaceticus, Pantoea agglomerans* and *ochrobactrum anthropic*). The isolates were cultured on Blood, MacConkey and Mannitol Salt agar

plates and incubated at 37°C for 24 hours. They were identified according to colony characteristics and microscopic examination of stained smear that demonstrate microbial shape, structure, agreement, gram stain reaction, and biochemical tests like indol (I), methyl red (M.R), vogas proskauer (V.P), citrate utilization, oxidase (C), motility test, catalase, coagulase, and urease production; according to (29) as in table (2). Then the identificatin was confirmed by using VITEK 02 compact system as recommended by Biomerieux. Our results were compared with the resource reported by  $(30)$  and  $(19)$ .

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*aeromonas anthropico Citrobacter freundii*





### **Antibacterial activities of Mgo NPs**

Antibacterial activity of 1.0 mg from Mgo NPs was tested against uncommon bacteria gram positive *Aeromonas salmonicida, Kocuria rosea and Alloiococcus otitis* and gram negative *Acinatobacter calcoaceticus, Pantoea agglomerans* and *ochrobactrum anthropic* bacteria. The results were indicated that highly inhibition activity of Mgo NPs against these bacterial pathogens and the zone inhibition diameter (ZID) against against Gram positive *Aeromonas salmonicida, Kocuria rosea and Alloiococcus otitis* and the zone inhibition diameter was at 20,19 and 23 mm respectively compared with inhibition effects of ciprofloxacin antibiotics at 18, 17 and 20 mm respectively. while against Gram negative *Acinatobacter calcoaceticus, Pantoea agglomerans* and *ochrobactrum anthropic* bacteria was at 18,20,19 mm respectively, compared with inhibition effects of ciprofloxacin antibi0tics at 16 , 20 and 15 mm respectively. Also The results demonstrated a slightly higher antibacterial activity against Gram positive bacteria than Gram negative bacteria these results were agreement with results founds by (26). Whom found that the amount of Mgo NPs which killed bacteria was strongly dependent on particle size, (31) said the gram positive bacteria more sensitive to oxides nanoparticles than the gram negative bacteria, The inhibition efficacy of Mgo NPs was tested against different pathogens (32)





The antibacterial activity effect of Mgo NPs on cell membrane integrity, which affects the permeability of membranes where nanoparticles enter and induce stress in

bacterial cells, subsequently resulting in the inhibition of cell growth and eventually in cell death (7, 33).

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*Acinatobacter calcoaceticus chrobactrum anthropico*

## **Fig. (4):- Antibacterial activity of Mgo NPs used well diffusion test**

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